

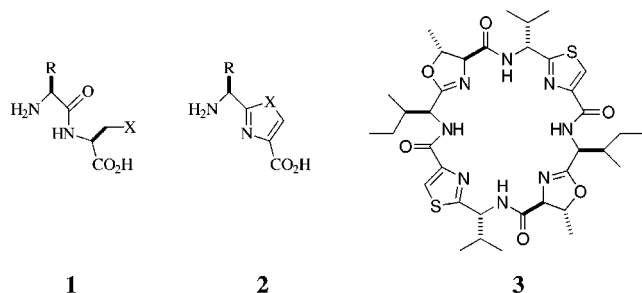
Novel Cylindrical, Conical, and Macrocyclic Peptides from the Cyclooligomerization of Functionalized Thiazole Amino Acids

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Intramolecular condensation of cysteine, serine, or threonine side chains of dipeptides Aaa-Cys, Aaa-Ser, or Aaa-Thr (**1**) results in dipeptide surrogates (**2**) that incorporate five-membered heterocyclic ring constraints such as thiazole (X = S), oxazole (X = O) or their reduced analogues. Such dipeptide surrogates have been found in many natural products, where they often profoundly influence three-dimensional structures and bioactivities.^{1–3} The number and oxidation state of heterocyclic rings can enforce macro-chair or macro-boat conformations on macrocycles such as **3**,⁴ and this has been shown to result in very different affinities for metal ions such as Cu²⁺,^{5a} Zn²⁺,^{5b} and Ca²⁺.^{5c} Dipeptide surrogates such as **2** have been exploited as β -turn-inducing constraints to control cyclooligomerizations⁶ used to form macrocycles such as [Ile-Ser-D-Val(Thz)]_{2–19}, with up to 76 amino acids in the cycle.^{6b}



We now report high-yielding cyclotrimerization reactions of functionalized dipeptide surrogates L-Glu(Thz) (**2a**, X = S, R =

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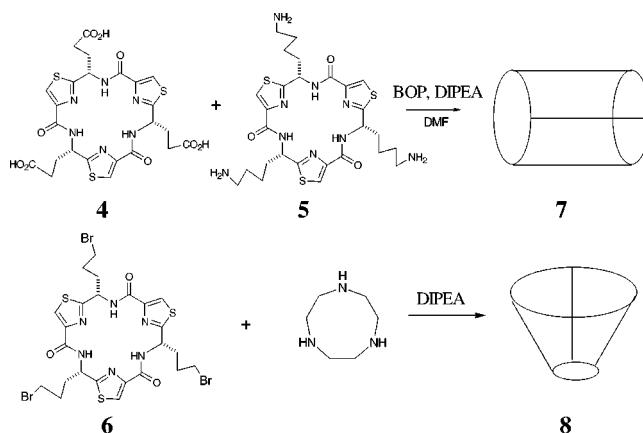
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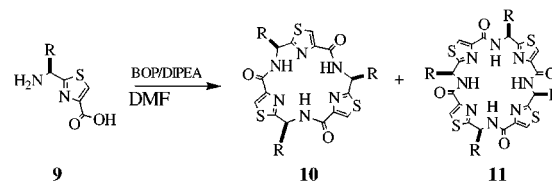
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–(CH₂)₂CO₂H) and L-Lys(Thz) (**2b**, X = S, R = –(CH₂)₄NH₂) into cyclic hexapeptides (**4–6**), and their subsequent elaboration to two new types of highly constrained chiral cavitands, the peptidic cylinder (**7**) and cone (**8**). Both the cylinder and cone are pentacyclic molecules, featuring two 18-membered macrocycles joined by three intercyclc linkages that create three additional 30-membered rings. These unique peptide cavitands illustrate new classes of supramolecular peptides for conceivable further development to catalysts and artificial proteins.



Cyclic hexapeptide **4** was prepared by converting the protected amino acid BocGlu(OcHx)-OH to the thiazole dipeptide BocGlu(OcHx)Thz-OtBu in 86% overall yield by an established method.⁷ This involved derivatization of BocGlu(OcHx)-OH to the primary amide, conversion to the corresponding thioamide with Lawesson's reagent, followed by a modified Hantzsch synthesis using *tert*-butyl bromopyruvate to give the protected thiazole amino acid. Treatment of the protected thiazole with TFA yielded **9** (R = –(CH₂)₂CO₂CHx). Cyclooligomerization of **9** (2 × 10^{–3} M) with BOP using DIPEA as base and DMF as solvent gave high isolated yields of cyclic hexapeptide **10a** (85%) and cyclic octapeptide **11a** (10%) where R = –(CH₂)₂CO₂CHx, which were separated by column chromatography.⁸ This contrasts with poor cyclooligomerization yields reported for similar oxazole-amino acids.⁹ Removal of the cyclohexyl-protecting group from **10a** with HF produced the cyclic hexapeptide **4**. Cyclooligomerization proceeds equally well at higher concentrations of **9** (85% **10a** at 4 × 10^{–2} M; 83% at 0.1 M).



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(8) Typically: TFA·H-Glu(OcHX)Thz-OH (3.5 g, 8.2 mmol) and BOP (5.5 g, 12.4 mmol) were dissolved in DMF (200 mL), cooled to 0 °C, DIPEA (10 mL, 57.4 mmol) added, and stirred (0 °C, 5 h). Solvent was evaporated in vacuo, and the residue was dissolved (150 mL EtOAc), washed with 20% aqueous citric acid (1 × 50 mL), saturated NaHCO₃ (4 × 60 mL), water (1 × 80 mL), and brine (50 mL), dried (anhydrous MgSO₄), and concentrated. Chromatography on Si gel with petroleum ether/EtOAc gave **10a** cyclo-[Glu(OcHx)Thz]₃ (2.0 g, 85%; HRMS: M + H exptl 1177.4253, calcd 1177.4230) and **11a** cyclo-[Glu(OcHx)Thz]₄ (241 mg, 10%; HRMS: M + H exptl 883.3201, calcd 883.3192).

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Cyclic peptide **5** was similarly prepared by sequential conversion of FmocLys(Boc)-OH via FmocLys(Boc)-NH₂ to FmocLys(Boc)Thz-OEt (96% overall yield), followed by selective deprotection of the ethyl ester and Fmoc groups. Surprisingly the Fmoc- and ethyl ester protecting groups were not easily removed with LiOH or KOH. These reagents cleanly removed the Fmoc group, while the ethyl ester was resistant to hydrolysis. Tetramethylammonium hydroxide however, did remove both protecting groups. Subsequent cyclooligomerization at 4×10^{-2} M and deprotection of the side chain with TFA yielded 95% cyclic products, as cyclic hexapeptide **5** (83%) and cyclic octapeptide **11b** (R = -(CH₂)₄-NH₂) (12%), which were readily separated by rpHPLC.

To investigate the generality and efficiency of this reaction, the alternatively protected dipeptide analogue H-Lys(Z)Thz-OH (**2c**, X = S, R = -(CH₂)₄NHCOCH₂C₆H₅) was cyclooligomerized at differing concentrations. Lower combined isolated yields of cyclic trimer **10b** plus tetramer **11b** (R = -(CH₂)₄NH-Z) were obtained (71% at 1×10^{-2} M, 56% at 0.1 M) due to competition from polymerization. H-Lys(Z)Thz-OH was similarly obtained from BocLys(Z)-OH as described above for H-Lys(Boc)Thz-OH. When an oxazole ring constraint (Ox) was present instead of thiazole, cyclooligomerization of H-Glu(OcHx)Ox-OH (**2d**, X = O, R = -(CH₂)₂CO₂C₆H₁₁) at 4×10^{-2} M gave respectable isolated yields of the oxazole analogues of **10a** and **11a**, namely cyclo-[Glu(OcHx)Ox]₃ (48%) and cyclo-[Glu(OcHx)Ox]₄ (16%), under conditions used for H-Glu(OcHx)Thz-OH (**9**).⁸

Cyclic peptides **4** and **5** are planar, by analogy with the crystal structure of c[Ala(Ox)D-Val(Thz)Gly(Thz)],¹⁰ and are rigid templates that direct their three L-Lys or L-Glu side chains from the same face of the macrocycle. ¹H NMR spectra for cyclic hexapeptides **4**, **5**, and **10** [R = -(CH₂)₂CO₂tBu or -(CH₂)₄NHBoc] gave only one set of resonances for their three dipeptide units, indicating high C₃ symmetry. In addition, the chemical shift (δ 8.43) of the amide-NH resonance of **10** [R = -(CH₂)₂CO₂tBu] is independent of temperature (CDCl₃, $\Delta\delta/T$ = 0.6 ppb/K) and does not significantly exchange in CD₃OD after 1 h. This is consistent with the amide-NH protons being involved in intramolecular H-bonding which would fix a planar conformation. The ¹H NMR spectrum for cyclic octapeptide **11** [R = -(CH₂)₂CO₂tBu] similarly indicates high C₄ symmetry. However, the chemical shift (δ 7.91 ppm) for the single amide-NH resonance of **11** is more temperature-dependent (CDCl₃, $\Delta\delta/T$ = 3 ppb/K) than for **10** [R = -(CH₂)₂CO₂tBu] and exchanges instantly in CD₃OD. Here the symmetry is due to conformational averaging not rigidity.

Cylinder **7** was synthesized by coupling cycle **4** with a stoichiometric equivalent of cycle **5** at 1×10^{-3} M using BOP, DIPEA, and DMF.¹¹ Some HMPA and hydroxybenzotriazole (degradation from BOP) were trapped by **7**, shown by their co-elution from rpHPLC columns and by ¹H NMR spectra, suggesting potential host-guest properties for **7**. These were removed slowly by stirring with NaHCO₃ in aqueous DMF. The cylinder was isolated pure in 47% yield, its ¹H NMR spectrum (Figure 1) is simple due to C₃ symmetry, and resonances for both sets of hexapeptide rings are distinct.

The cone-shaped molecule **8** was prepared by reduction of the macrocyclic triacid **4** by activation with BOP followed by reduction with NaBH₄ to the tri-alcohol (83%). This was converted

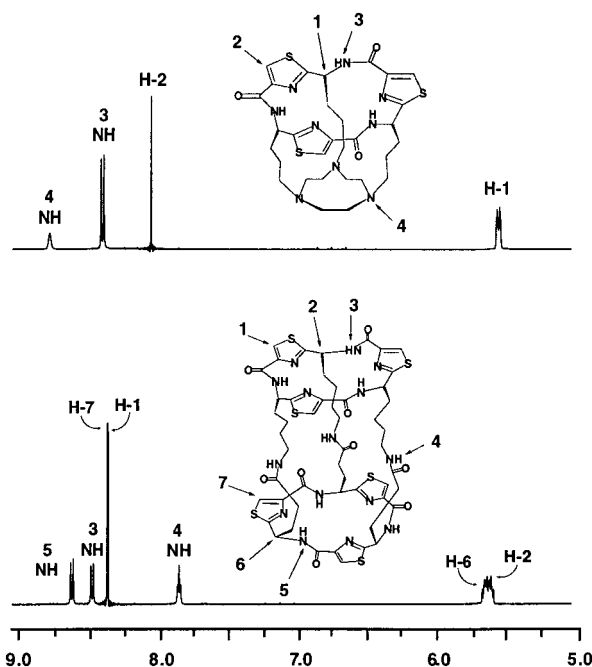


Figure 1. ¹H NMR spectra (5–9 ppm) for **7** (bottom) and **8** (top) in *d*₆-DMSO at 293 K.

with CBr₄/PPh₃ to the tribromide **6** in 57% yield. Reaction of **6** with 1,4,7-triazacyclononane (TACN) in DMF (0.002 M) with DIPEA as base gave the cone **8** (55%).¹² The ¹H NMR spectrum (Figure 1) is consistent with C₃ symmetry, although interestingly only one of the three TACN nitrogens appears to be protonated based on proton integration. The CH_α-CH_β coupling constants for the tribromide **6** ($J_{\text{H}\alpha\text{-H}\beta}$ = 5.7 Hz) are significantly larger than for the cone **8** ($J_{\text{H}\alpha\text{-H}\beta}$ = 3.9 Hz, $J_{\text{H}\alpha\text{-H}\beta}$ = 2.8 Hz), consistent with a smaller H_α-C-C-H_β dihedral angle in **8** and more conformational restriction. Preliminary CD studies suggest that **8** interacts with Cu²⁺ in MeOH indicating potential host-guest properties for **8**.

This work has demonstrated that highly functionalized thiazole-amino acids can be cyclooligomerized in high yields to constrained cyclic hexa- and octa-peptides. These cycles have been used as scaffolds to construct unique conical and cylindrical peptides, which can potentially capture small molecules, metal ions, and short peptides within their interiors. In cones of this type all three nitrogen lone pairs project into the interior¹³ due to the high barrier to inversion in TACN. Modified cylinders, enlarged through the use of bigger cycles and extended through insertion of amino acids or peptides between Glu and Lys side chains, could conceivably be developed as mimics of pseudocylindrical enzyme active sites. This new approach to cavitands promises to lead to novel classes of molecular receptors.

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Supporting Information Available: Syntheses and characterization (NMR, MS, HPLC) for all compounds (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(11) Cyclo-(Glu(Thz))-**4** (106 mg, 0.17 mmol), cyclo-(Lys(Thz))-**3**TFA **5** (163 mg, 0.17 mmol) and BOP reagent (320 mg, 0.72 mmol) were dissolved in DMF (160 mL), DIPEA (0.60 mL, 3.44 mmol) was added, and the mixture was stirred (65 h, 20 °C). After evaporation of the solvent in vacuo, the residue was stirred with concentrated NaHCO₃ for 30 h, freeze-dried, and purified by HPLC to yield **7** (95 mg, 47%; HRMS: M + H exptl 670.2437, calcd 670.2416).

(12) Cyclo-(Glu(CH₂Br)(Thz))-**3** **6** (79 mg, 0.1 mmol) was dissolved in DMF (50 mL) and DIPEA (0.18 mL, 1 mmol) to which was added 1,4,7-triazacyclononane (2 mL of 0.1 M solution in dioxane). The mixture was stirred overnight at 50 °C and evaporated to a residue that was purified by preparative HPLC to yield **8** (43 mg, 55%; HRMS: M + H exptl 1216.2912, calcd 1216.2866).

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